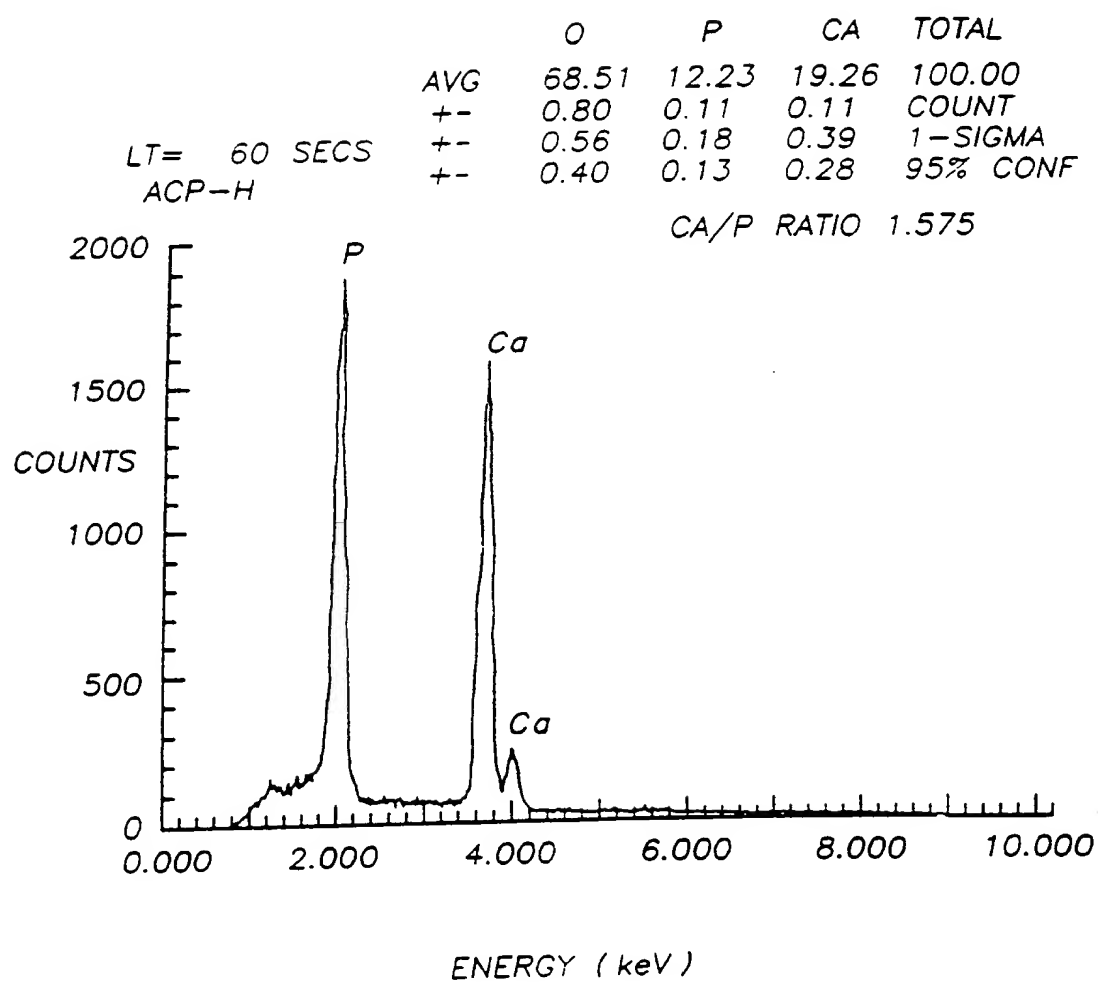


FIG. 1

09/284436

FIG. 2



09/284436

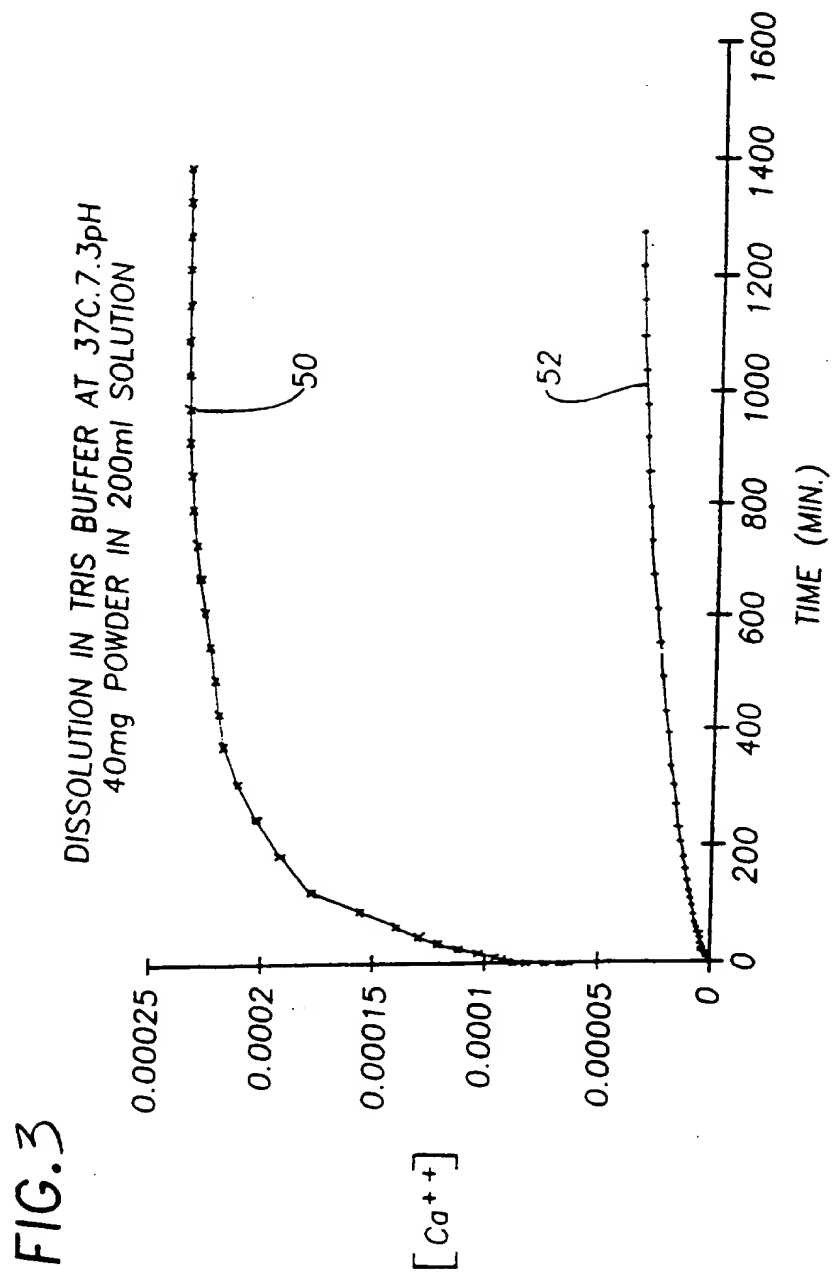
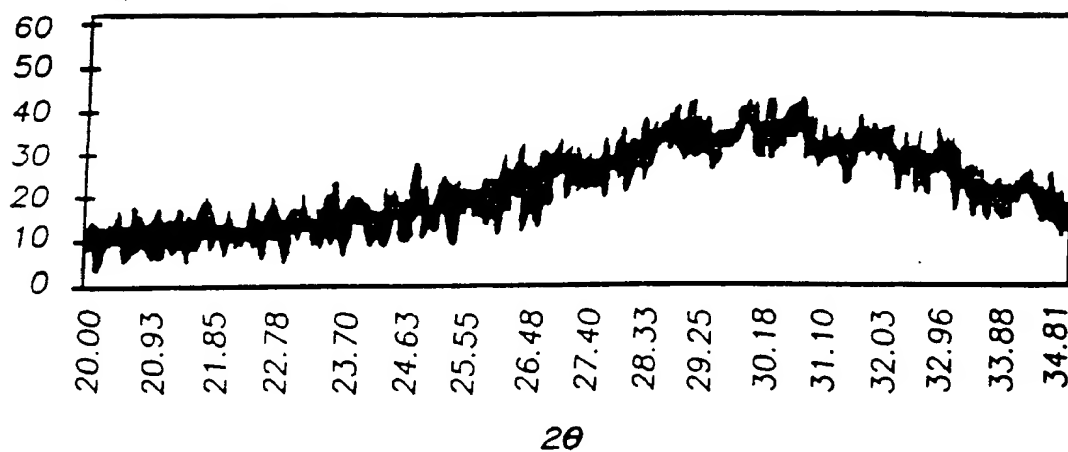


FIG. 4

(a)

REACTIVE ACP

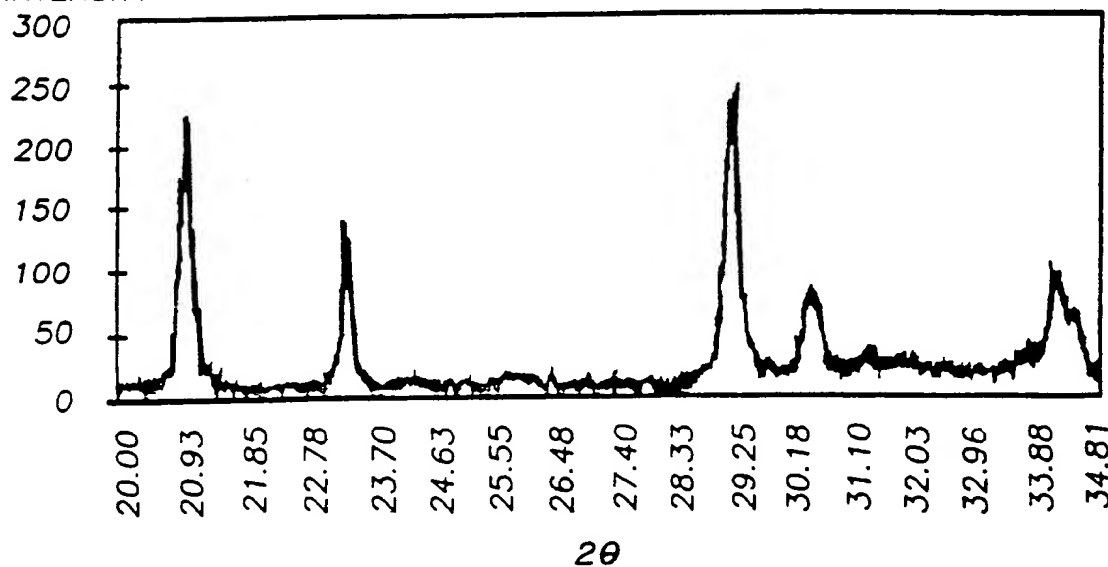
INTENSITY



(b)

DCDP

INTENSITY



09/284436

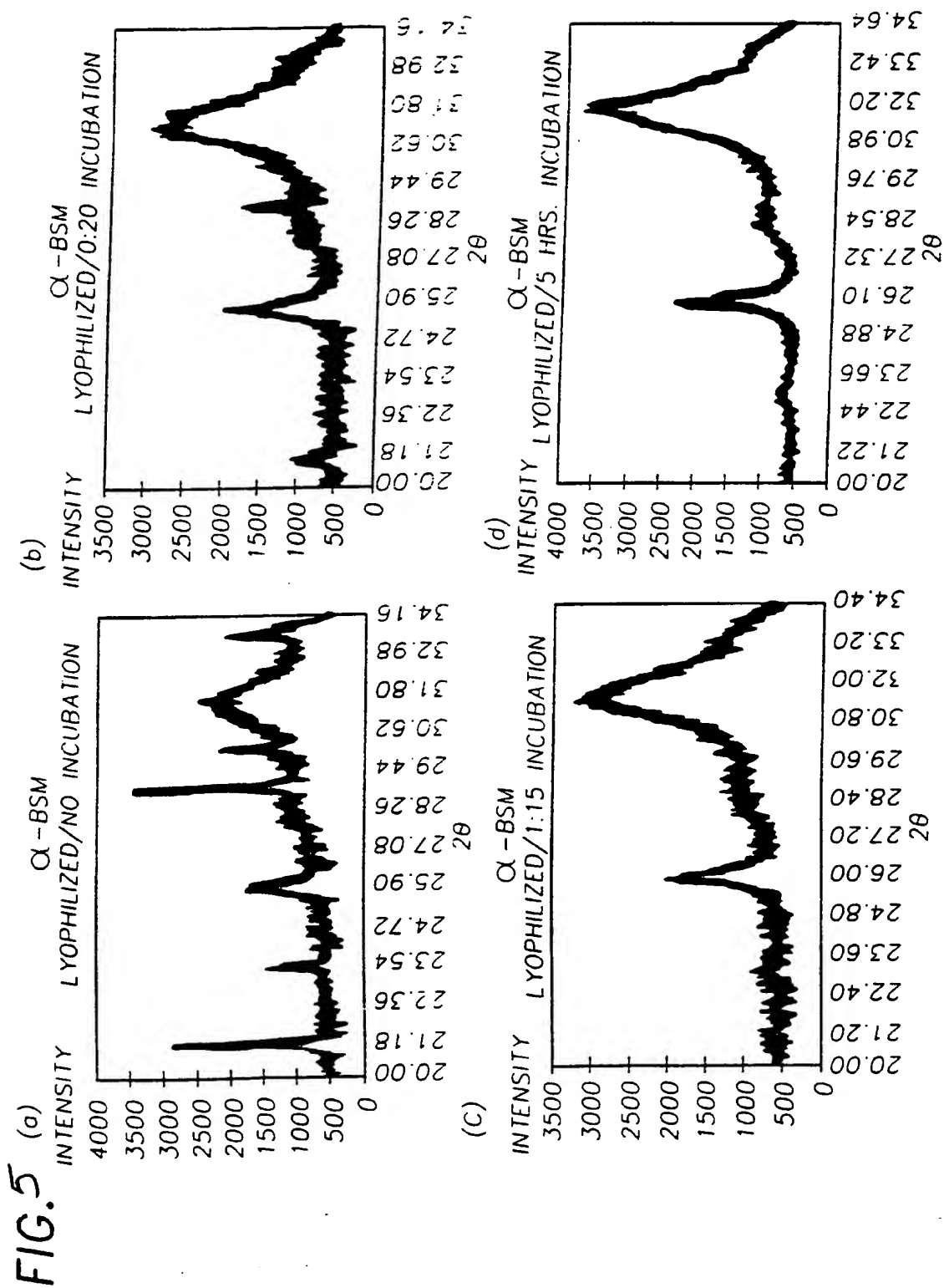


FIG. 3

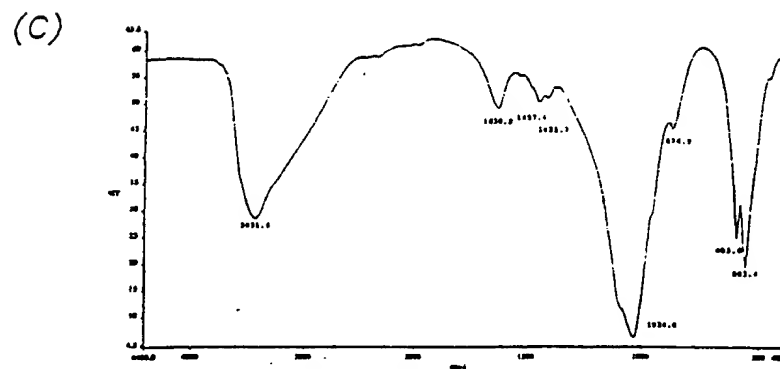
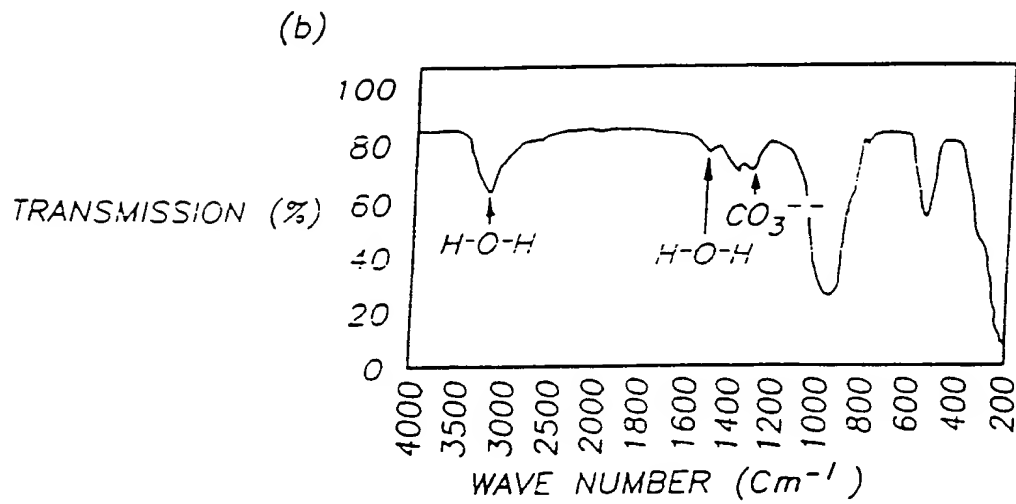
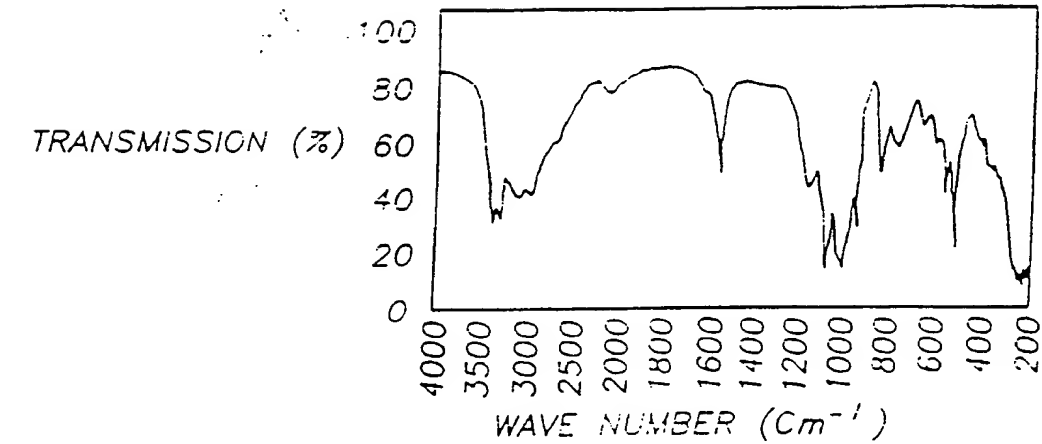
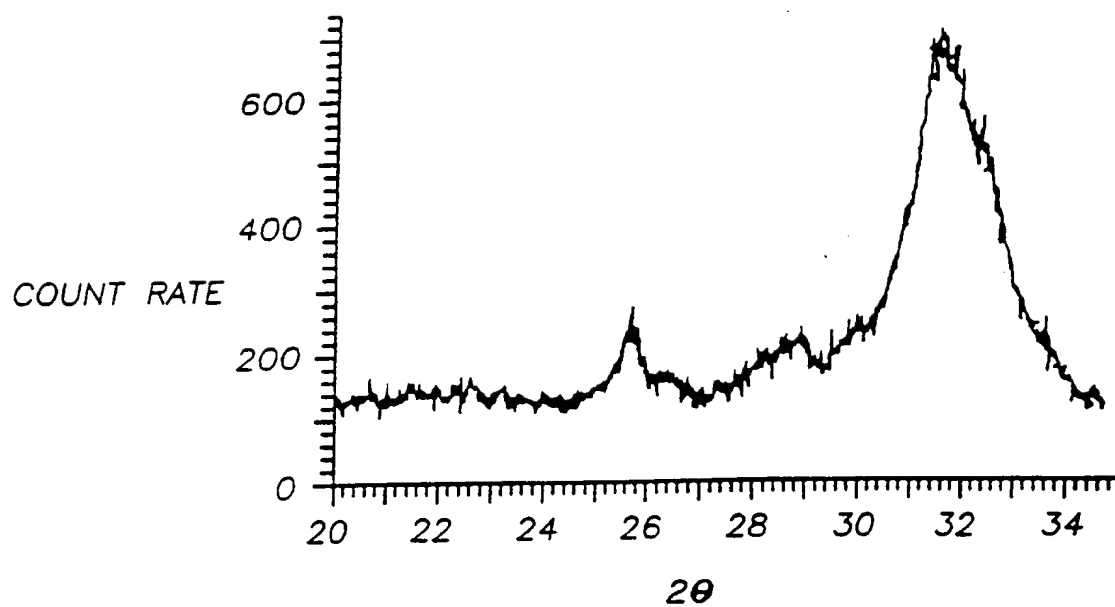


FIG. 7



% of powder 'B' Vs. Particle Size

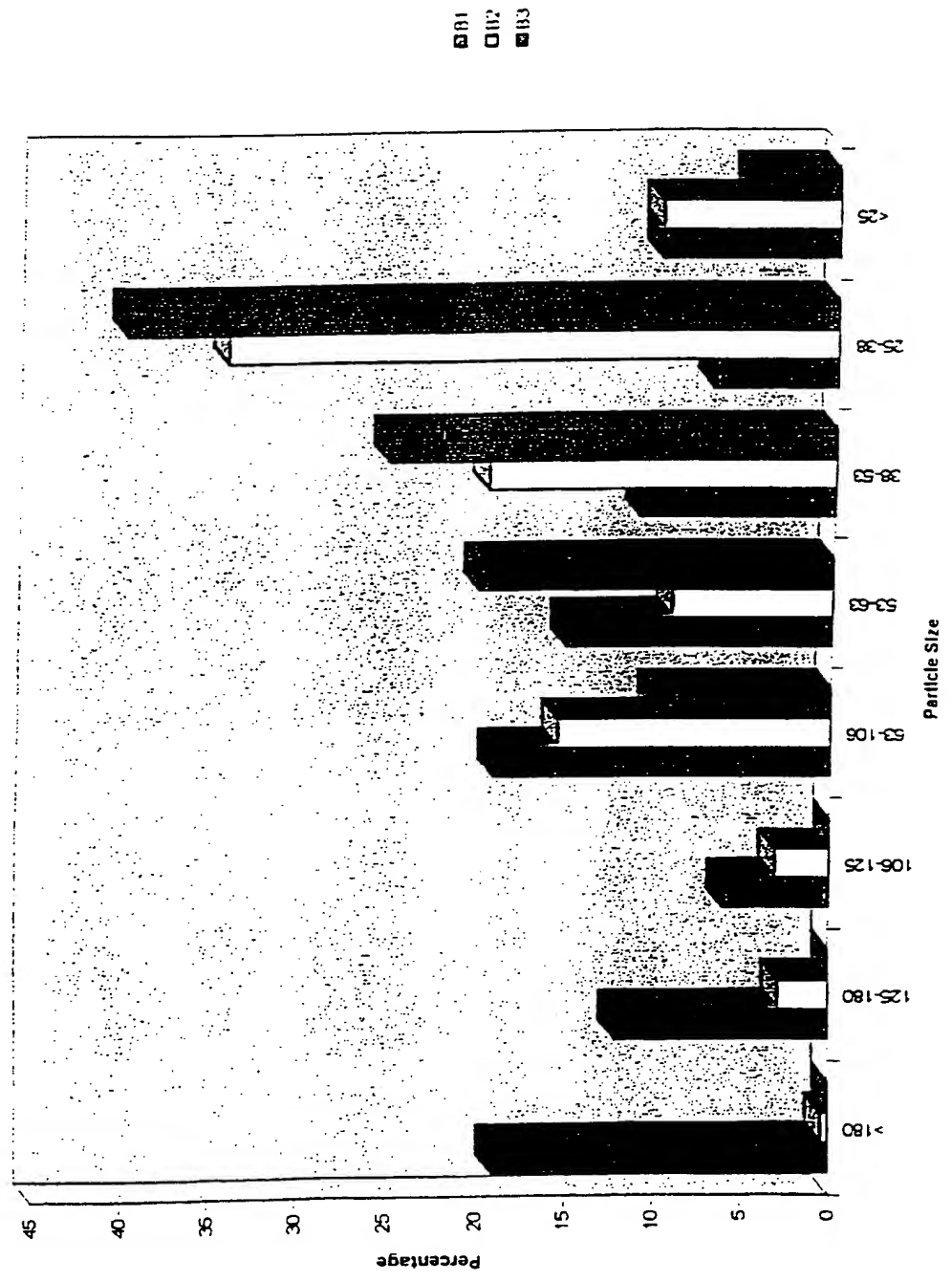


FIG 8

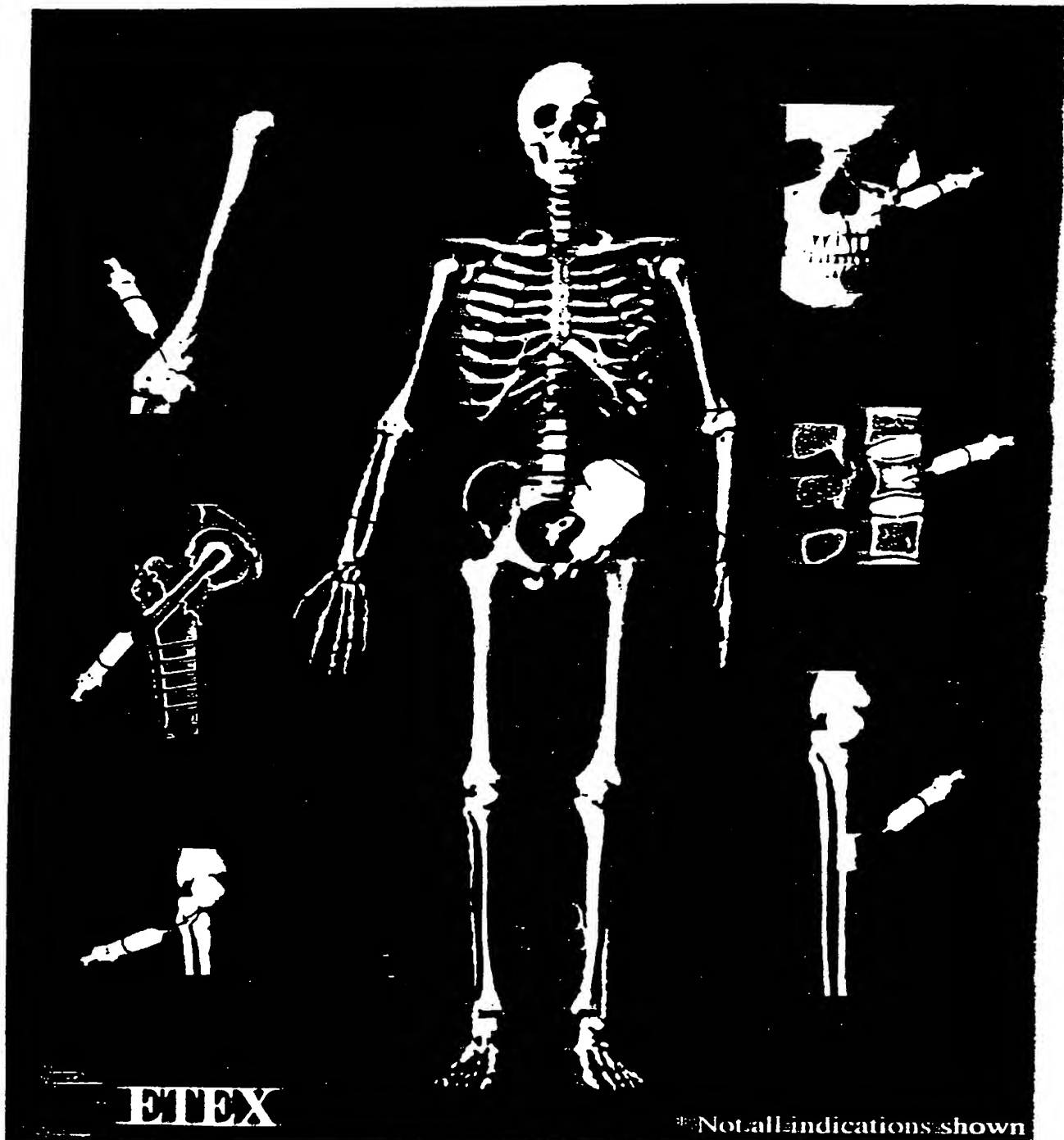


FIG 9

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09/284436

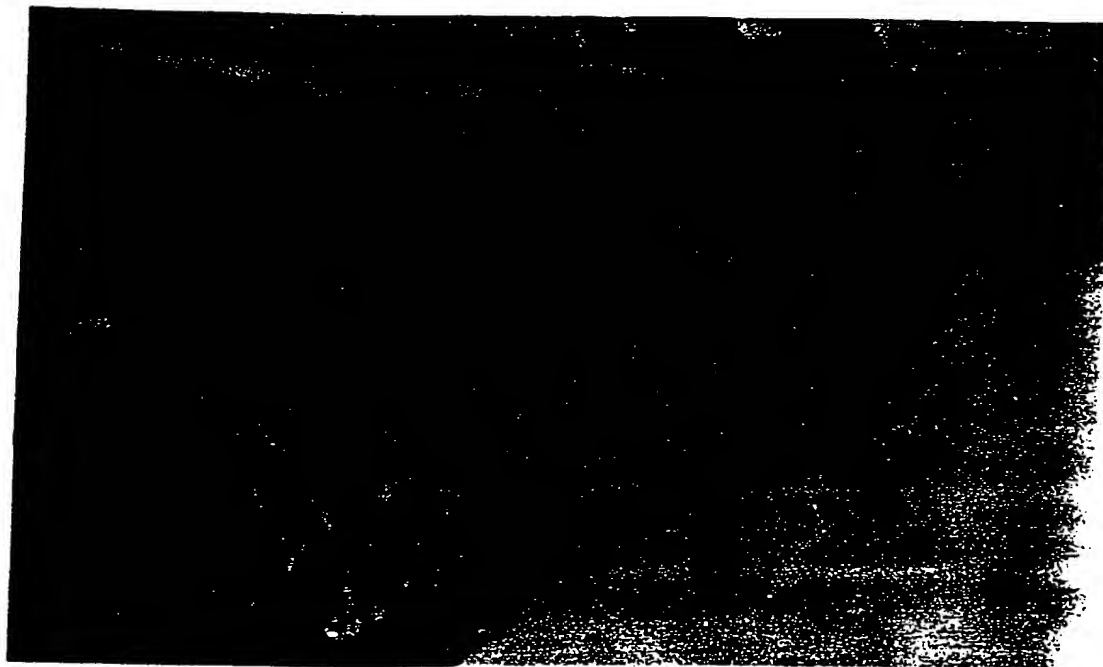
WO 98/16209

10/22

PCT/US97/18528

Study EX96-1-002

Bone Substitute Material (BSM™) Screening Assay in the NZW Rabbit Proximal
Tibia Bone Defect Model



Photomicrograph of untreated control rabbit #72 to a defect 2 weeks after surgery. The small arrows indicate one edge of the created defect. The large arrowhead is at the yet unbridged defect. Bone present to the right of the defect edge is trabecular bone. Magnification 4x decalcified, hematoxylin & eosin.

Fig 10a

Study EX96-1-002
Bone Substitute Material (BSM™) Screening Assay in the NZW Rabbit Proximal
Tibia Bone Defect Model



Photomicrograph of a bone defect treated with BSM from rabbit #712 weeks after surgery. Large arrowheads denote one edge of the defect. New bone to the right of the defect edge is thick trabecular bone. Magnification 4X, decalcified, hematoxylin and Eosin.

FIG. 10D

Study EX95-1-004

Pilot Efficacy Study of Bone Substitute Material (BSM™) in the Canine Proximal Tibia Bone Defect Model



Photomicrograph of canine trabecular bone grown into the defect site treated with BSM. The small arrows denote osteoblast-like cells lining the bone spicules and are indicative of enhanced cellular activity. (Magnification 10X; decalcified; Hematoxylin & Eosin).

FIG. 11

Study EX95-1-004**Pilot Efficacy Study of Bone Substitute Material (BSM™) in the Canine Proximal Tibia Bone Defect Model**

Photomicrograph of a canine cortical bone defect that was treated with BSM. The large arrows indicate one edge of the defect. The new bone growth is to the right of the defect and at 4 weeks after surgery is thick trabecular bone. (Magnification 4X, undecalcified, Light Green Basic Fuchsin)

FIG. 12

Study EX95-1-005

Establishment of a Bone Substitute Material (BSM™) Screening Assay in the NZW Rabbit Proximal Tibia Bone Defect Model

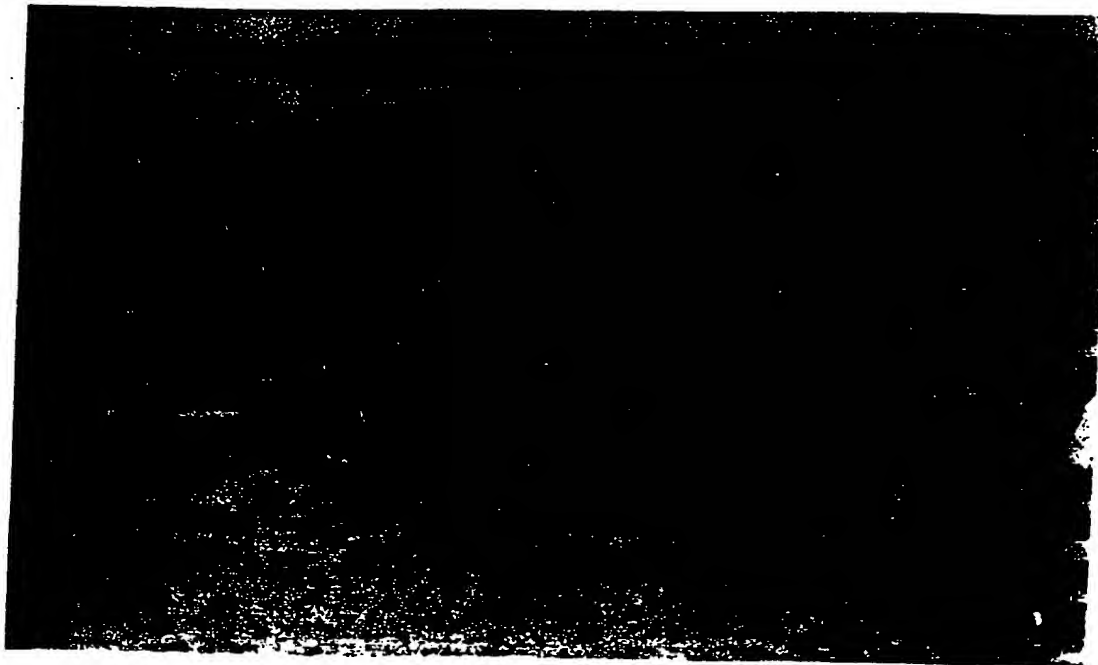


Photomicrograph of an untreated (control) tibia defect in rabbit #31 at 4 weeks after surgery. The large arrow indicates the edge of the defect. The small arrowheads indicate the remaining defect with no bone. Small arrows denote an abundance of fibrous connective tissue in the defect site. The large arrowhead points to new trabecular bone in the defect. (Magnification 4X, decalcified Masson's Trichrome.)

FIG. 13a

Study EX95-1-005

Establishment of a Bone Substitute Material (BSM) Screening Assay in the NZW Rabbit Proximal Tibia Bone Defect Model



Photomicrograph of a bone defect from rabbit #41 treated with BSM at 4 weeks after surgery. The large arrowheads delineate the edge of the defect. The 2 small arrows demarcate the trabecular bone grown into the defect site. Magnification 4X. Decalcified, Hematoxylin & Eosin.

Fig. 13b

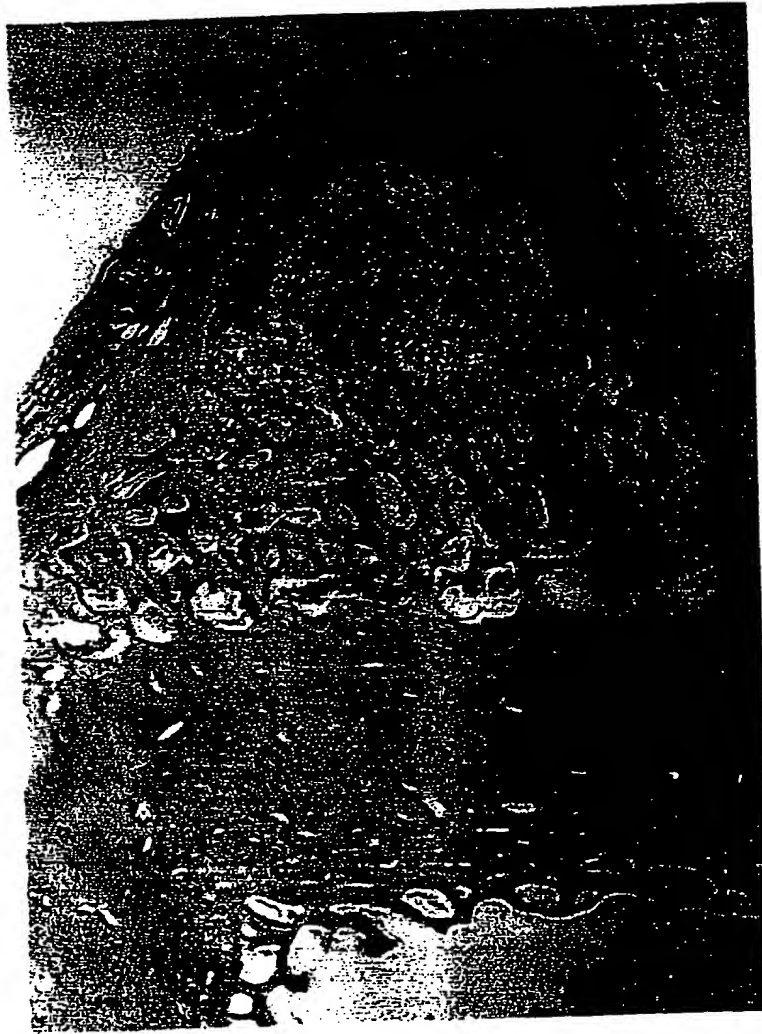


FIG. 14

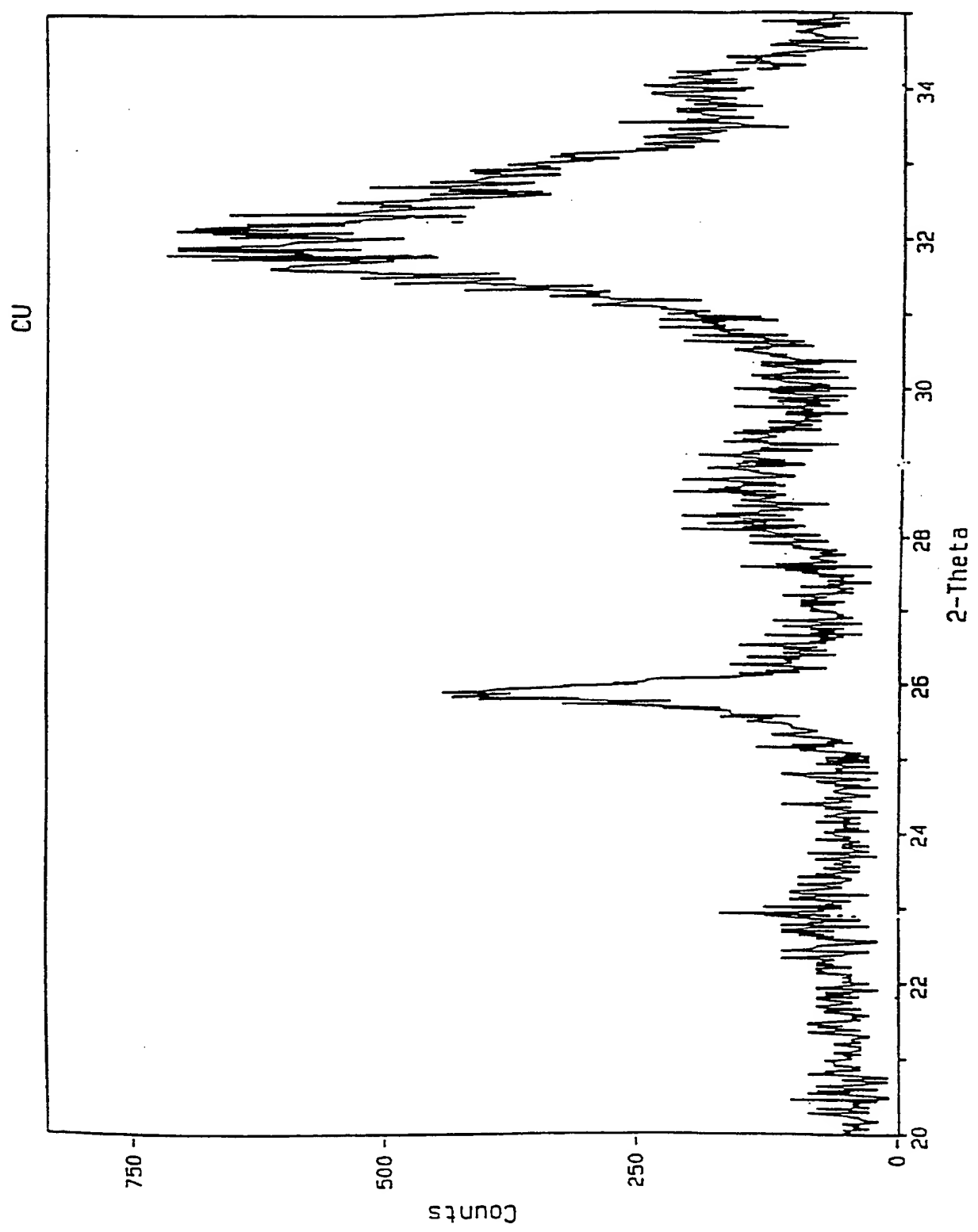


Figure 15

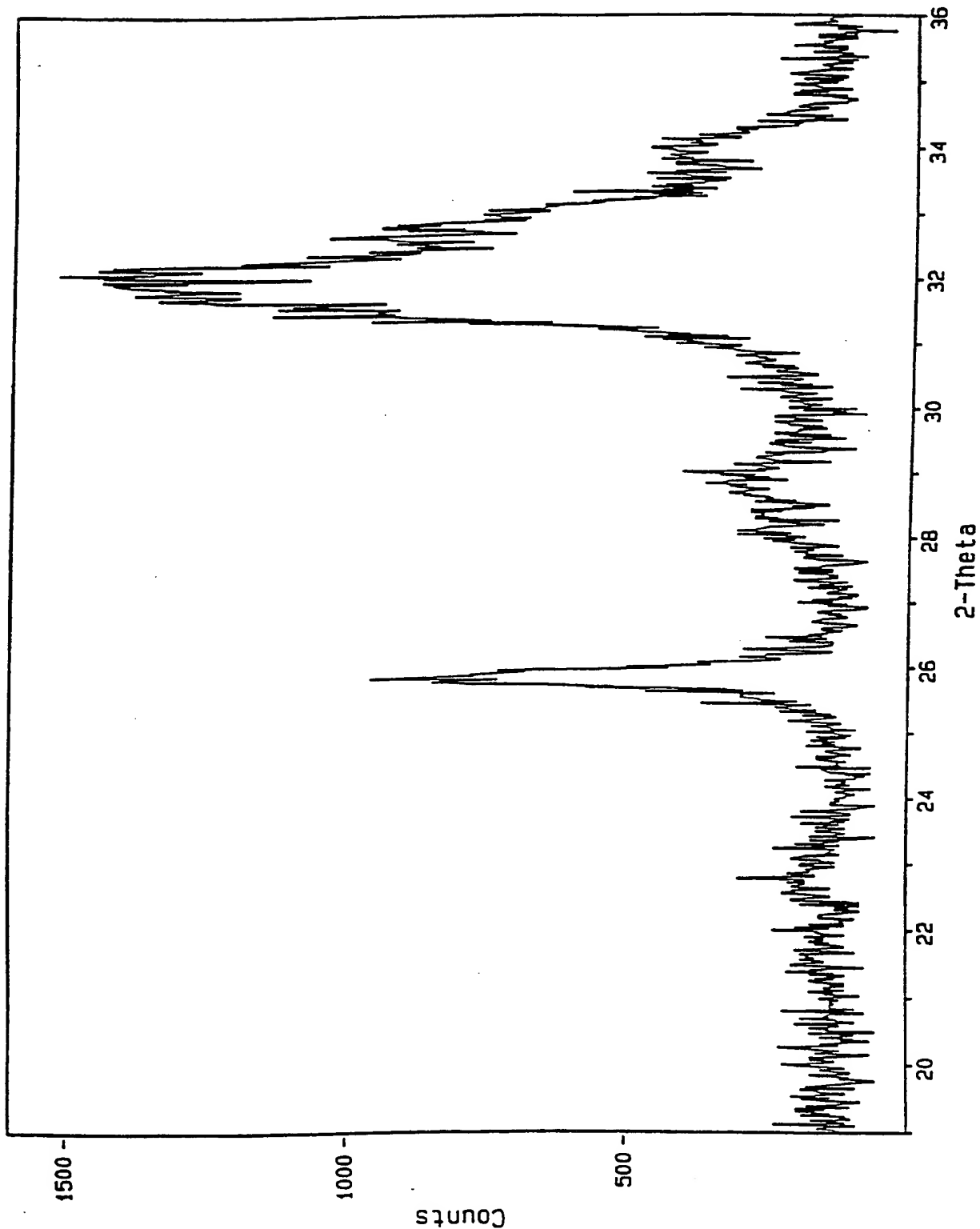


Figure 16

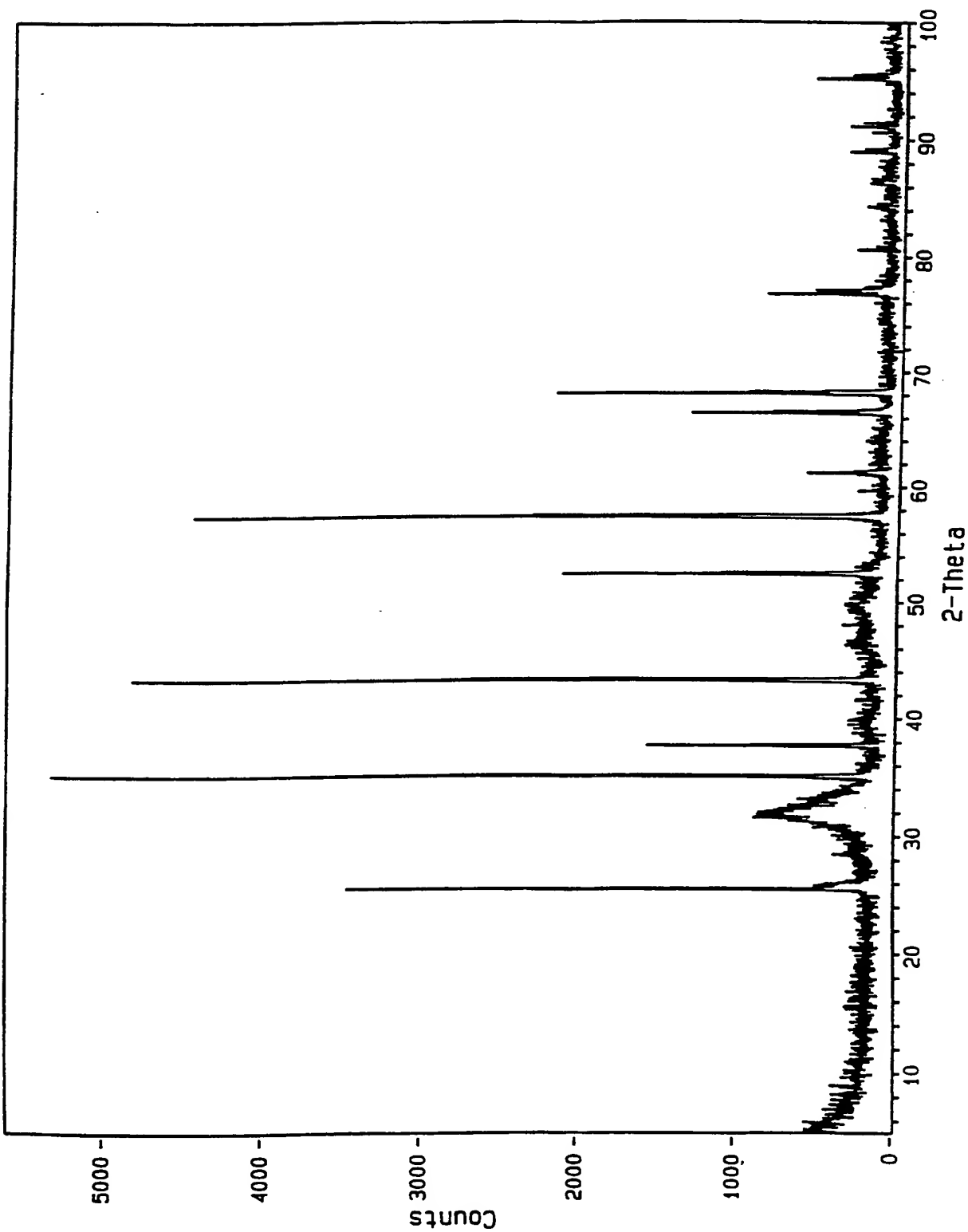
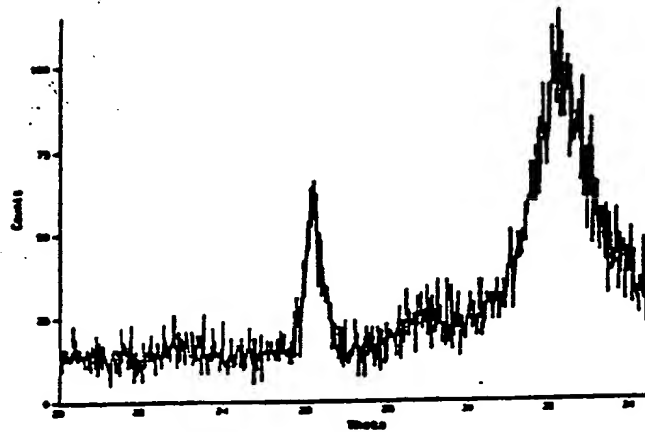


Figure 17

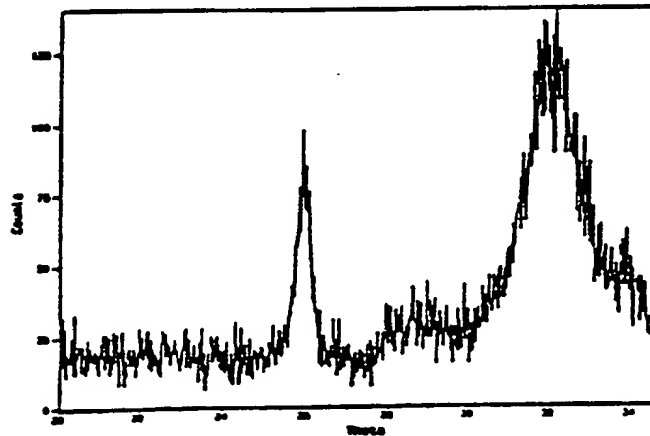
ETEX C . Confidential Report
Study 96-008

XRD ANALYSIS OF EXPLANTED α -BSM™ FOR DAYS 4,7,14

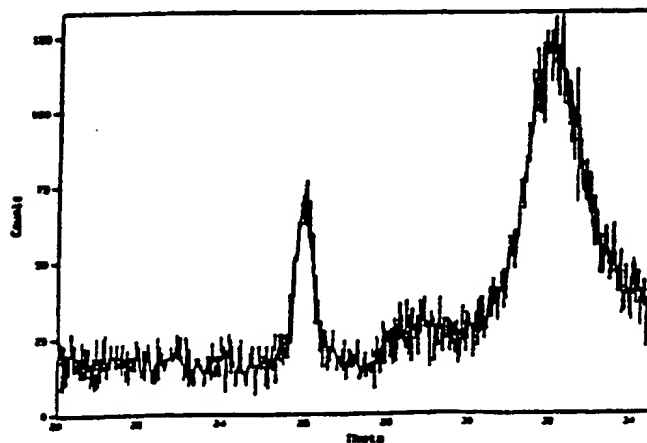
Panel 1



Panel 2

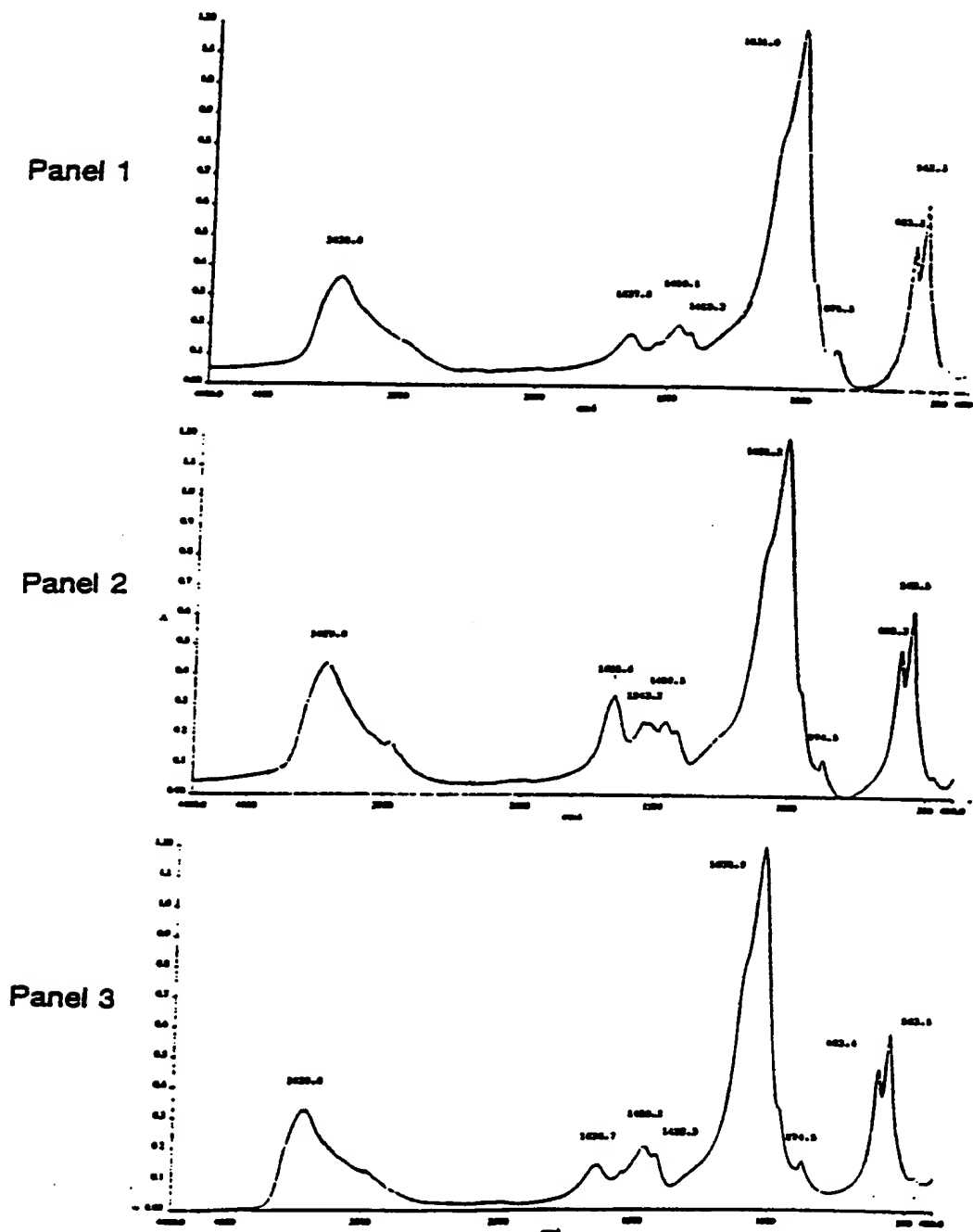


Panel 3

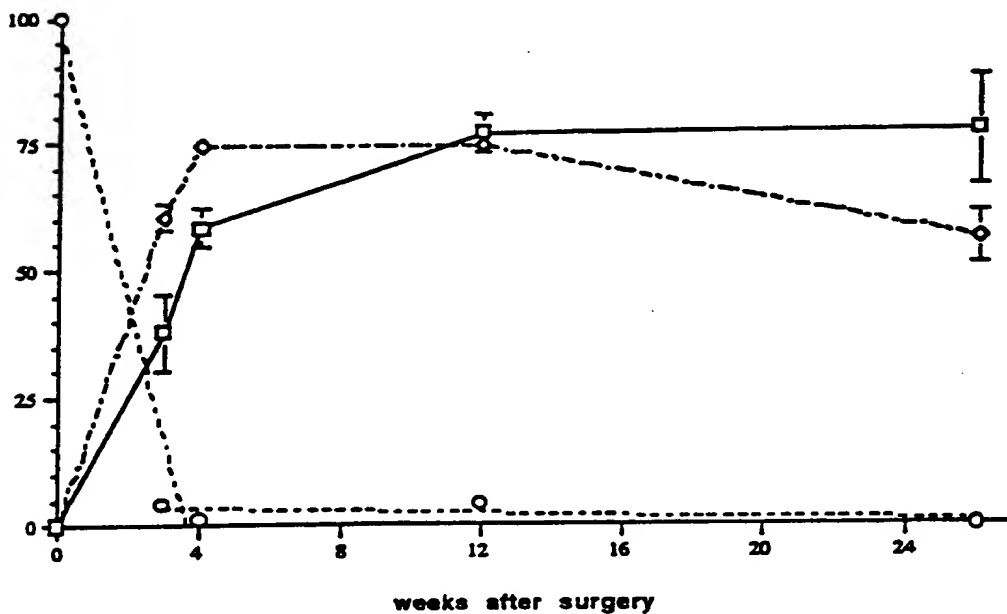


ETEX C Confidential Report
Study 96-08

FTIR ANALYSIS OF EXPLANTED α -BSM™ FOR DAYS 4,7,14



α -BSM™ Resorption and Defect Healing Compared to Autograft Healing



This figure demonstrates the resorption of α -BSM™ (circles) following implantation into a canine femoral defect. Also represented is the formation of new bone within the defect site, for animals treated with either α -BSM™ (squares) or with autologous bone (diamonds). The data is presented as the % of the defect occupied by calcium phosphate (either new bone or α -BSM™) as determined by light microscope histomorphometry of von Kossa stained undecalcified sections. Error bars represent standard error of the mean. For weeks 3 and 26, n=4; For weeks 4 and 12, n=8

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